

# ACR–SPR–SSR PRACTICE PARAMETER FOR THE PERFORMANCE AND INTERPRETATION OF MAGNETIC RESONANCE IMAGING (MRI) OF BONE, JOINT, AND SOFT-TISSUE INFECTIONS IN THE EXTREMITIES

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## PREAMBLE

This document is an educational tool designed to assist practitioners in providing appropriate radiologic care for patients. Practice Parameters and Technical Standards are not inflexible rules or requirements of practice and are not intended, nor should they be used, to establish a legal standard of care<sup>1</sup>. For these reasons and those set forth below, the American College of Radiology and our collaborating medical specialty societies caution against the use of these documents in litigation in which the clinical decisions of a practitioner are called into question.

The ultimate judgment regarding the propriety of any specific procedure or course of action must be made by the practitioner considering all the circumstances presented. Thus, an approach that differs from the guidance in this document, standing alone, does not necessarily imply that the approach was below the standard of care. To the contrary, a conscientious practitioner may responsibly adopt a course of action different from that set forth in this document when, in the reasonable judgment of the practitioner, such course of action is indicated by variables such as the condition of the patient, limitations of available resources, or advances in knowledge or technology after publication of this document. However, a practitioner who employs an approach substantially different from the guidance in this document may consider documenting in the patient record information sufficient to explain the approach taken.

The practice of medicine involves the science, and the art of dealing with the prevention, diagnosis, alleviation, and treatment of disease. The variety and complexity of human conditions make it impossible to always reach the most appropriate diagnosis or to predict with certainty a particular response to treatment. Therefore, it should be recognized that adherence to the guidance in this document will not assure an accurate diagnosis or a successful outcome. All that should be expected is that the practitioner will follow a reasonable course of action based on current knowledge, available resources, and the needs of the patient to deliver effective and safe medical care. The purpose of this document is to assist practitioners in achieving this objective.

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<sup>1</sup> *Iowa Medical Society and Iowa Society of Anesthesiologists v. Iowa Board of Nursing*, 831 N.W.2d 826 (Iowa 2013) Iowa Supreme Court refuses to find that the "ACR Technical Standard for Management of the Use of Radiation in Fluoroscopic Procedures (Revised 2008)" sets a national standard for who may perform fluoroscopic procedures in light of the standard's stated purpose that ACR standards are educational tools and not intended to establish a legal standard of care. See also, *Stanley v. McCarver*, 63 P.3d 1076 (Ariz. App. 2003) where in a concurring opinion the Court stated that "published standards or guidelines of specialty medical organizations are useful in determining the duty owed or the standard of care applicable in a given situation" even though ACR standards themselves do

*not establish the standard of care.*

## I. INTRODUCTION

This practice parameter was developed and written collaboratively by the American College of Radiology (ACR), the Society for Pediatric Radiology (SPR), and the Society of Skeletal Radiology (SSR).

Bone, joint, and soft-tissue infections of the extremities are challenging conditions for clinicians and radiologists. Efficient diagnosis and timely treatment are important to prevent long-term morbidity. Evaluation of the patient with suspected musculoskeletal (MSK) infection affecting an extremity combines clinical assessment, laboratory investigations, and diagnostic imaging.

Osteomyelitis (bone infection) [1-3], septic arthritis (infection of a joint) [1,3-5], and deep and superficial soft-tissue infections [6-16] occur in all age groups. They are caused by a variety of bacteria and less commonly by viruses, fungi, or parasites [1-3,12,17]. The routes of contamination include hematogenous, direct inoculation, and contiguous spread [1,3,6-8,10,13,14,16].

Magnetic resonance imaging (MRI) has proven to be one of the most useful imaging modalities in the evaluation of MSK infections [1,7,8,10,20-24]. In the extremities, MRI is usually the study of choice when it is necessary to confirm or exclude clinically suspected infections, to stage the local extent of disease, and to follow-up on patients after treatment. Compared with other imaging modalities, the power of MRI is its sensitivity for detecting bone marrow abnormalities and its ability to characterize associated soft-tissue abnormalities [3,21]. Because early diagnosis and treatment of infection leads to a better prognosis and more efficient treatment, MRI should be done early if septic arthritis and/or osteomyelitis is suspected [25-28]. For osteomyelitis, the negative predictive value of an appropriately performed MRI approximates 100%; a normal study practically excludes active infection [20,22].

Radiography [8,21,29-32], computed tomography (CT) [6,22,33-40], ultrasound [21,22], and combined bone and labeled leukocyte scintigraphy [41-46] have complementary roles in the evaluation of MSK infections. Radiography should be the initial imaging test. It may demonstrate findings of established osteomyelitis, or, in the case of some soft-tissue infections, may reveal gas or foreign matter. Initial radiographs may show an alternative diagnosis, such as fracture or tumor that accounts for the clinical symptoms and obviates further evaluation for infection. Additionally, radiographs help the interpretation of MRI studies, especially in the diabetic, neuropathic, or postoperative foot, in which infection is often superimposed on neuropathic disease and surgically altered anatomy [22,31]. Nuclear medicine examinations, including bone scintigraphy and labeled leukocyte scans, have a potential role in the detection of infection in specific circumstances, especially with multifocal osteomyelitis or when infection is suspected near metallic implants or superimposed on pre-existing bone disease [41,42,47-50]. The introduction of single-photon PET (SPECT)/CT imaging has provided improved diagnostic accuracy over that of planar or SPECT-alone scans. The main value of SPECT/CT is more of precise anatomical localization of infection and accurate delineation of the infection extent after its diagnosis with planar scintigraphy [51]. Additionally, in patients with contraindications precluding MRI, nuclear medicine imaging may be used for primary diagnosis. Fluorine-18-2-fluoro-2-deoxy-D-glucose positron emission tomography (FDG-PET) has also shown promising results in identifying MSK infections in specific situations [1,22,52-58]. The FDG-PET test is sensitive, with a high negative predictive value, and it reliably differentiates degenerative from infectious vertebral body end plate abnormalities [59].

Compared with modalities other than ultrasound, which is primarily used to evaluate for soft-tissue abscesses, MRI does not use ionizing radiation and provides superb evaluation of the bone marrow and soft tissues [1,22]. CT provides better visualization of cortical bone than MRI [33] and may better depict a sequestrum. However, the viability of infected bone in acute and subacute infection and the presence of intraosseous abscesses are better defined using MRI [33,60]. Although MRI and ultrasound can be used to detect gas in necrotizing infections, CT is the most sensitive modality and can usually be performed faster than MRI for critically ill patients [61,62]. Although CT is highly sensitive for the detection of gas, the absence of gas does not exclude necrotizing fasciitis [63-65]. Patients with contraindications to MRI will require other modalities for primary evaluation. Additionally, although not a contraindication, metallic implants distort the magnetic field, and the resulting artifacts degrade

image quality. Therefore, imaging parameters should be optimized using metal artifact reduction (MAR) techniques or advanced MAR sequences should be used if available [66,67]. In selected cases, use of >1 imaging modality will be needed for a complete evaluation [68-71]. Furthermore, CT and ultrasound play a role in guiding aspiration and biopsy of infected bones, joints, and soft tissues [22,33].

Despite its strengths, MRI should be performed only for a valid medical reason, and its findings need to be interpreted in conjunction with clinical history, physical examination, and laboratory results to avoid misinterpretations [16,72]. Adherence to the following practice parameter will increase the probability of detecting clinically relevant abnormalities in patients with bone, joint, and soft-tissue infections in the extremities.

## II. INDICATIONS

Indications for MRI of bone, joint, and soft-tissue infections of the extremities include, but are not limited to, screening, staging, and follow-up of:

1. Bone infection including, but not limited to:
  - a. Acute osteomyelitis [1,22]
  - b. Subacute osteomyelitis [1,22]
  - c. Chronic osteomyelitis [1,22]
  - d. Complications of osteomyelitis [3,28,73]
2. Septic arthritis and its complications [1,22]
3. Soft-tissue infections including, but not limited to:
  - a. Cellulitis refractory to initial treatment [16,20,35,37,74,75]
  - b. Superficial fasciitis [16,21,75]
  - c. Deep fasciitis, including necrotizing fasciitis [16,64,75,76]
  - d. Soft-tissue abscess and/or pyomyositis [16,26,75,77]
  - e. Septic tenosynovitis [16,21,78]
  - f. Septic bursitis [16,21]
  - g. Infectious lymphadenitis [6,16]
  - h. Deep and superficial septic thrombophlebitis [79]
  - i. Complications of soft-tissue infections [79]

## III. QUALIFICATIONS AND RESPONSIBILITIES OF PERSONNEL

See the [ACR Practice Parameter for Performing and Interpreting Magnetic Resonance Imaging \(MRI\)](#) [80].

## IV. SAFETY GUIDELINES AND POSSIBLE CONTRAINDICATIONS

See the [ACR Practice Parameter for Performing and Interpreting Magnetic Resonance Imaging \(MRI\)](#) [80] and the [ACR Guidance Document on MR Safe Practices 2020](#) [81].

Peer-reviewed literature pertaining to MR safety should be reviewed on a regular basis [82,83].

## V. SPECIFICATIONS OF THE EXAMINATION

The written or electronic request for MRI of bone, joint and soft tissue infections of the extremities should provide sufficient information to demonstrate the medical necessity of the examination and allow for its proper performance and interpretation.

Documentation that satisfies medical necessity includes 1) signs and symptoms and/or 2) relevant history (including known diagnoses). Additional information regarding the specific reason for the examination or a provisional diagnosis would be helpful and may at times be needed to allow for the proper performance and interpretation of the examination.

The request for the examination must be originated by a physician or other appropriately licensed health care provider. The accompanying clinical information should be provided by a physician or other appropriately licensed health care provider

familiar with the patient's clinical problem or question and consistent with the state's scope of practice requirements. (ACR Resolution 35 adopted in 2006 – revised in 2016, Resolution 12-b)

The supervising physician must understand the indications, risks, and benefits of the imaging examination, as well as alternative imaging procedures. The physician must be familiar with potential hazards associated with MRI, including potential adverse reactions to contrast media. The physician should be familiar with relevant ancillary studies that the patient may have undergone. The physician performing MRI interpretation must have a clear understanding and knowledge of the anatomy and pathophysiology relevant to the MRI examination.

The supervising physician must also understand the pulse sequences to be used and their effect on the appearance of the images, including the potential generation of image artifacts. Standard imaging protocols may be established and varied on a case-by-case basis when necessary. These protocols should be reviewed and updated periodically.

#### A. Patient Selection

The physician responsible for the examination should supervise patient selection and preparation and be available in person or by phone for consultation. Patients must be screened and interviewed prior to the examination to exclude individuals who may have contraindications to MRI, in which the risks may outweigh the benefits.

Certain indications require administration of intravenous (IV) contrast media. IV contrast enhancement should be performed using appropriate injection protocols and in accordance with the institution's policy on IV contrast utilization (see the [ACR–SPR Practice Parameter for the Use of Intravascular Contrast Media \[84\]](#)).

Pediatric patients or patients suffering from anxiety or claustrophobia may require sedation or additional assistance. Administration of sedation or general anesthesia may be needed to achieve a successful examination, particularly in young children. If minimal or moderate sedation is necessary, refer to the [ACR–SIR Practice Parameter for Minimal and/or Moderate Sedation/Analgesia \[85\]](#).

#### B. Facility Requirements

Appropriate emergency equipment and medications must be immediately available to treat adverse reactions associated with administered medications. The equipment and medications should be monitored for inventory and drug expiration dates on a regular basis. The equipment, medications, and other emergency support must also be appropriate for the range of ages and sizes in the patient population.

#### C. Examination Technique

Diagnostic-quality MRI of suspected bone, joint, and soft-tissue infections of the extremities can be performed using a variety of magnet designs (closed-bore whole body, open-bore whole body, dedicated extremity) and a variety of field strengths [8,70,86-92]. Regardless of system design, efforts should be made to maximize signal-to-noise (SNR) ratios [93]. Field of view (FOV) should be tailored to the size of the patient and the size of the suspected abnormality [94,95]. For example, a 48-cm FOV may be appropriate for evaluating a suspected large focus of infection in the pelvis or thigh, whereas a 12-cm or smaller FOV may be appropriate for a suspected focal infection in a finger or toe. At times, additional sequences with a larger FOV will be necessary to evaluate proximal or distal spread of disease. An initial survey sequence with a large FOV is also appropriate in infants and young children because of the difficulty in precisely localizing sites of involvement by clinical examination and the frequent multifocality of involvement in this population [96,97]. Slice thicknesses also will vary depending on the size of the lesion. For example, a small, infected focus might require 3-mm-thick slices or thinner, whereas an infection that involves the majority of one extremity may be appropriately imaged with 8-mm slice thickness. The imaging matrix should balance intravoxel SNR with desired in-plane spatial resolution.

The size of the lesion and desired spatial resolution will also dictate the choice of coil, which might be a local surface or cylindrical coil for a small lesion or a multicoil array to completely image a more extensive

area (eg, the entire lower extremity for suspected necrotizing fasciitis). Ideally, the size of the coil will closely match the image volume, balancing anatomic coverage with optimal SNR. For patients with suspected multifocal infection, it may be necessary to perform separate MRI examinations of the affected parts of the extremities, each using a separate coil. For example, a patient with infection involving both the hip and hand will require 2 separate studies.

When using a low-field MRI system for the extremities, other imaging parameters—such as the receiver bandwidth and number of acquisitions—will require modification to ensure adequate spatial and contrast resolution for confident diagnosis, often at the expense of longer examination times [95,98]. It may be more difficult to achieve uniform fat suppression on low-field systems using spectrally selective radiofrequency (RF) presaturation pulses, potentially necessitating the use of Dixon or short-tau inversion recovery (STIR) techniques [99-102]. Additionally, specific systems may be more prone to artifacts (eg, chemical shift artifact on high-field magnets), again requiring that parameters like receiver bandwidth be optimized to ensure that these artifacts do not detract from the diagnostic quality of the resultant images. Finally, some MRI systems may not be appropriate for specific indications. For example, high-resolution evaluation of a small focal lesion in a digit may not be feasible with a low-field open magnet, regardless of the chosen imaging parameters [103].

The examination should include images in both short and long axes. The long-axis images may be oriented orthogonal to the magnetic bore or may be angled to better identify specific anatomic structures. The coverage of the lesion ideally should include the entire infection focus [95,104] plus as much of the surrounding inflammation as is reasonably feasible.

MRI of extremity infections can be performed with a variety of pulse sequences. The choice of sequences may be tailored to optimize the examination for specific clinical questions and according to local preferences. In general, however, fast spin-echo (FSE) or turbo spin-echo (TSE) images are preferred [94,95,104]. Gradient-recalled sequences may also be valuable, particularly in evaluating for internal areas of hemorrhage, gas, ossification, foreign material, or calcification [95]. Gradient-echo images, however, are relatively insensitive to changes in marrow composition and would need to be supplemented by other sequences when evaluating for osteomyelitis. Imaging sequences using isotropic or near-isotropic 3-D sequences can produce images with shorter scan duration, but have not been evaluated for imaging extremity MSK infections [105]. For any chosen sequence, the exact recovery time (TR), echo time (TE), and flip angle used will depend on the field strength of the magnet and the relative contrast weighting desired. An imaging protocol for a MSK infection typically will comprise >1 pulse sequence type but should include, at a minimum, a fluid-sensitive sequence. A T1-weighted sequence is useful for bone infections. The planes of imaging will depend on reader preference and the region being imaged. Although the water-sensitive images are the most sensitive for areas of marrow and soft-tissue edema, they may overestimate the amount of osteomyelitis; the extent of infected bone (as opposed to reactive bone) is more accurately determined with T1-weighted sequences [106-108].

In many cases it is advantageous to administer a gadolinium-based IV contrast agent to increase conspicuity of infected tissues and to depict rim enhancement in intraosseous and soft-tissue abscesses. Typically, T1-weighted fat-suppressed sequences are obtained before and after contrast administration [16,21]. Subtraction of nonfat suppressed precontrast and postcontrast images may also be helpful to highlight enhancement when fat suppression techniques are likely to fail, such as in patients with metallic hardware [109]. Additionally, isolated infection within the completely or predominantly cartilaginous epiphyses of infants and younger children may be only conspicuous on postcontrast images, appearing as hypoenhancing or nonenhancing foci within the cartilage [18,19]. The slice orientation on the contrast-enhanced images depends on the imaged regional anatomy but is usually in the short-axis plane; many practices obtain additional contrast-enhanced images in a second (long-axis) plane [21,22,93,110]. In addition to showing areas of enhancement, detecting nonenhancement in infected bones and soft tissues impacts management because these nonviable tissues may require surgical debridement [74]. The decision to use IV contrast should be based on medical appropriateness (see the [ACR–SPR Practice Parameter for the Use of Intravascular Contrast Media](#) [84]) and should be undertaken only after consideration of potential adverse reactions (see the [ACR Manual on Contrast Media](#) [111]).

More advanced imaging techniques, such as diffusion-weighted imaging (DWI), dynamic contrast-enhanced (DCE) MRI, MR spectroscopy and PET-MR can be used in the evaluation of bone, joint, and soft-tissue infections as well; however, their role is currently being defined [24,112,113]. Diffusion-weighted imaging has been shown to be comparable with contrast-enhanced MRI in detection of soft-tissue abscesses and thus can be used when gadolinium-based IV contrast is contraindicated [77].

Various techniques may be used to minimize artifacts that can reduce image quality [114]. Wraparound artifact, including that which originated from signal received from other parts of the body, can be reduced by phase oversampling by switching the phase and frequency readout directions, by presaturation pulses, or by radiofrequency shielding. Achieving uniform T1 and T2 spectral fat suppression when imaging the hand/wrist and foot/ankle is often challenging. Current Dixon techniques often provide superior fat suppression in these locations [115]. Involuntary patient motion is best controlled by ensuring patient comfort combined with gentle immobilization when necessary [95,116]. The use of high field-strength systems and multichannel coil or coil array may allow the use of parallel imaging, compressed sensing, and machine-learning acceleration techniques to reduce overall scan time and/or improve SNR, and may be useful in reducing motion-related artifacts [95,104,117,118]. These acceleration techniques may, however, introduce new artifacts or change the appearance of conventional artifacts, although many of these artifacts can be minimized by optimizing protocols. It is important to have an understanding of the benefits and limitations of these techniques and when they should and should not be performed [119,120]. Flowing blood can produce ghosting artifacts, which can be reduced with presaturation pulses or the use of gradient moment nulling.

Artifacts also occur at interfaces between structures with different magnetic susceptibilities, especially where ferromagnetic materials are present in the body. Common examples include vascular filters, dental restorations, and orthopedic implants [121]. Techniques that can reduce metal artifacts include positioning of the patient with the long axis of instrumentation parallel to the main magnetic field, using fast spin-echo sequences with relatively long echo train lengths and short interecho spacing, substituting inversion recovery for chemical fat suppression, controlling phase and frequency encoding direction, employing view angle tilting, increasing bandwidth during slice selection or readout, and decreasing voxel size [122,123]. Specific metal artifact-reducing sequences, such as slice encoding for metal artifact correction (SEMAC) and multiacquisition variable-resonance image combination (MAVRIC) provide both in and through plane distortion corrections [123-126]. Susceptibility artifacts from surgical implants are more prevalent at higher field strengths (3T), and patients with known metallic implants are often scheduled at lower field strengths (1.5T) [127] [128].

MRI examinations in patients with suspected extremity infections should be interpreted in conjunction with all available clinical data and relevant imaging studies, including current radiographs, when possible. Inflammatory, metabolic, and neoplastic conditions can mimic infections based on their MRI appearances alone. For example, rheumatoid or gouty arthritis may be impossible to distinguish from septic arthritis on MRI [31]. It may also be difficult to distinguish soft-tissue abscesses from diabetic myonecrosis, necrotic soft-tissue tumors, and posttraumatic or postoperative seromas [16]. The signal intensity of reactive marrow edema (eg, in neuropathic arthropathy) can mimic that of osteomyelitis and can enhance with intravenous contrast agents, thus causing false-positive results [130]. Furthermore, imaging artifacts also can contribute to incorrect staging/evaluation of MSK infections [121].

## VI. DOCUMENTATION

Reporting should be in accordance with the [ACR Practice Parameter for Communication of Diagnostic Imaging Findings](#) [131].

The report should address the presence or absence of a bone, joint, or soft-tissue infection, the extent of the infection, and enhancement characteristics. A description of the anatomic location of a lesion, including its relationships to adjacent bone, joint, and soft-tissue structures (including the skin and neurovascular bundles) should be provided. The presence or absence of any regional lymphadenopathy should be noted. Other coexistent MSK abnormalities, especially those that may impact treatment planning, should also be recorded.

Specific policies and procedures related to MRI safety should be in place with documentation that is updated annually and compiled under the supervision and direction of the supervising MRI physician and, if available, MRI physicist. Guidelines that deal with potential hazards associated with MRI examination to the patient as well as to others in the immediate area should be provided [82,83,132,133]. Screening forms must also be provided to detect those patients who may be at risk of adverse events associated with the MRI examination [82,83,132,133].

## VII. EQUIPMENT SPECIFICATIONS

Equipment monitoring should be in accordance with the [ACR–AAPM Technical Standard for Diagnostic Medical Physics Performance Monitoring of Magnetic Resonance \(MR\) Imaging Equipment \[134\]](#).

The MRI equipment specifications and performance must meet all state and federal requirements. The requirements include, but are not limited to, specifications of maximum static magnetic strength, maximum rate of change of the magnetic field strength (dB/dt), maximum radiofrequency power deposition (specific absorption rate), and maximum acoustic noise levels.

### VIII. QUALITY CONTROL, AND IMPROVEMENT, SAFETY, INFECTION CONTROL, AND PATIENT EDUCATION

Policies and procedures related to quality, patient education, infection control, and safety should be developed and implemented in accordance with the ACR Policy on Quality Control and Improvement, Safety, Infection Control, and Patient Education appearing under the heading *ACR Position Statement on Quality Control & Improvement, Safety, Infection Control, and Patient Education* on the ACR website (<https://www.acr.org/Advocacy-and-Economics/ACR-Position-Statements/Quality-Control-and-Improvement>).

### IX. ACKNOWLEDGEMENTS

This practice parameter was developed according to the process described under the heading *The Process for Developing ACR Practice Parameters and Technical Standards* on the ACR website (<https://www.acr.org/Clinical-Resources/Practice-Parameters-and-Technical-Standards>) by the Committee on Practice Parameters – Body Imaging (Musculoskeletal) of the ACR Commission on Body Imaging and the Committee on Practice Parameters – Pediatric Radiology of the ACR Commission on Pediatric Radiology, in collaboration with the SPR and the SSR

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## REFERENCES

1. Christian S, Kraas J, Conway WF. Musculoskeletal infections. *Seminars in roentgenology* 2007;42:92-101.
2. Kothari NA, Pelchovitz DJ, Meyer JS. Imaging of musculoskeletal infections. *Radiologic clinics of North America* 2001;39:653-71.
3. Resnick D, Kransdorf M. *Bone and Joint Imaging*. 3rd ed. Philadelphia, Pa: Saunders; 2004.
4. Luhmann JD, Luhmann SJ. Etiology of septic arthritis in children: an update for the 1990s. *Pediatr Emerg Care* 1999;15:40-2.
5. Malleon PN. Management of childhood arthritis. Part 1: Acute arthritis. *Arch Dis Child* 1997;76:460-2.
6. Beauchamp NJ, Jr., Scott WW, Jr., Gottlieb LM, Fishman EK. CT evaluation of soft tissue and muscle infection and inflammation: a systematic compartmental approach. *Skeletal Radiol* 1995;24:317-24.
7. Beltran J. MR imaging of soft-tissue infection. *Magnetic resonance imaging clinics of North America* 1995;3:743-51.
8. Beltran J, Noto AM, McGhee RB, Freedy RM, McCalla MS. Infections of the musculoskeletal system: high-field-strength MR imaging. *Radiology* 1987;164:449-54.
9. Bosshardt TL, Henderson VJ, Organ CH, Jr. Necrotizing soft-tissue infections. *Arch Surg* 1996;131:846-52; discussion 52-4.
10. Brothers TE, Tagge DU, Stutley JE, Conway WF, Del Schutte H, Jr., Byrne TK. Magnetic resonance imaging differentiates between necrotizing and non-necrotizing fasciitis of the lower extremity. *J Am Coll Surg* 1998;187:416-21.
11. Chauhan S, Jain S, Varma S, Chauhan SS. Tropical pyomyositis (myositis tropicans): current perspective. *Postgrad Med J* 2004;80:267-70.
12. Elliott D, Kufera JA, Myers RA. The microbiology of necrotizing soft tissue infections. *Am J Surg* 2000;179:361-6.
13. Headley AJ. Necrotizing soft tissue infections: a primary care review. *Am Fam Physician* 2003;68:323-8.
14. Hill MK, Sanders CV. Necrotizing and gangrenous soft tissue infections. In: Sanders CV, Nesbitt LT, Jr, ed. *The*

*skin and infection: a color atlas and text*. Baltimore, Md: Williams & Wilkins; 1995:62-75.

15. McHenry CR, Piotrowski JJ, Petrinic D, Malangoni MA. Determinants of mortality for necrotizing soft-tissue infections. *Ann Surg* 1995;221:558-63; discussion 63-5.
16. Turecki MB, Taljanovic MS, Stubbs AY, et al. Imaging of musculoskeletal soft tissue infections. *Skeletal Radiol* 2010;39:957-71.
17. Savarese LG, Monsignore LM, de Andrade Hernandez M, Martinez R, Nogueira-Barbosa MH. Magnetic resonance imaging findings of paracoccidioidomycosis in the musculoskeletal system. *Trop Med Int Health* 2015;20:1346-54.
18. Johnson DP, Hernanz-Schulman M, Martus JE, Lovejoy SA, Yu C, Kan JH. Significance of epiphyseal cartilage enhancement defects in pediatric osteomyelitis identified by MRI with surgical correlation. *Pediatr Radiol* 2011;41:355-61.
19. Browne LP, Guillerman RP, Orth RC, Patel J, Mason EO, Kaplan SL. Community-acquired staphylococcal musculoskeletal infection in infants and young children: necessity of contrast-enhanced MRI for the diagnosis of growth cartilage involvement. *AJR Am J Roentgenol* 2012;198:194-9.
20. Gylys-Morin VM. MR imaging of pediatric musculoskeletal inflammatory and infectious disorders. *Magnetic resonance imaging clinics of North America* 1998;6:537-59.
21. Lalam RK, Cassar-Pullicino VN, Tins BJ. Magnetic resonance imaging of appendicular musculoskeletal infection. *Top Magn Reson Imaging* 2007;18:177-91.
22. Palestro CJ, Love C, Miller TT. Infection and musculoskeletal conditions: Imaging of musculoskeletal infections. *Best Pract Res Clin Rheumatol* 2006;20:1197-218.
23. Gilbertson-Dahdal D, Wright JE, Krupinski E, McCurdy WE, Taljanovic MS. Transphyseal involvement of pyogenic osteomyelitis is considerably more common than classically taught. *AJR Am J Roentgenol* 2014;203:190-5.
24. Pugmire BS, Shailam R, Gee MS. Role of MRI in the diagnosis and treatment of osteomyelitis in pediatric patients. *World J Radiol* 2014;6:530-7.
25. Gottschalk HP, Moor MA, Muhamad AR, Wenger DR, Yaszay B. Improving diagnostic efficiency: analysis of pelvic MRI versus emergency hip aspiration for suspected hip sepsis. *J Pediatr Orthop* 2014;34:300-6.
26. Mignemi ME, Menge TJ, Cole HA, et al. Epidemiology, diagnosis, and treatment of pericapsular pyomyositis of the hip in children. *J Pediatr Orthop* 2014;34:316-25.
27. Monsalve J, Kan JH, Schallert EK, Bisset GS, Zhang W, Rosenfeld SB. Septic arthritis in children: frequency of coexisting unsuspected osteomyelitis and implications on imaging work-up and management. *AJR Am J Roentgenol* 2015;204:1289-95.
28. Johnston JJ, Murray-Krezan C, Dehority W. Suppurative complications of acute hematogenous osteomyelitis in children. *J Pediatr Orthop B* 2017;26:491-96.
29. Manaster BJ, May DA, Disler DG *Musculoskeletal Imaging: The Requisites*. 2nd ed. St. Louis, MO: Mosby; 2002.
30. Mellado Santos JM. Diagnostic imaging of pediatric hematogenous osteomyelitis: lessons learned from a multi-modality approach. *Eur Radiol* 2006;16:2109-19.
31. Taljanovic MS, Hunter TB, Fitzpatrick KA, Krupinski EA, Pope TL. Musculoskeletal magnetic resonance imaging: importance of radiography. *Skeletal Radiol* 2003;32:403-11.
32. Vartanians VM, Karchmer AW, Giurini JM, Rosenthal DI. Is there a role for imaging in the management of patients with diabetic foot? *Skeletal Radiol* 2009;38:633-6.
33. Fayad LM, Carrino JA, Fishman EK. Musculoskeletal infection: role of CT in the emergency department. *Radiographics* 2007;27:1723-36.
34. Gordon BA, Martinez S, Collins AJ. Pyomyositis: characteristics at CT and MR imaging. *Radiology* 1995;197:279-86.
35. Ma LD, Frassica FJ, Bluemke DA, Fishman EK. CT and MRI evaluation of musculoskeletal infection. *Crit Rev Diagn Imaging* 1997;38:535-68.
36. Pretorius ES, Fishman EK. Helical CT of musculoskeletal infection. *Crit Rev Diagn Imaging* 2001;42:259-305.
37. Struk DW, Munk PL, Lee MJ, Ho SG, Worsley DF. Imaging of soft tissue infections. *Radiologic clinics of North America* 2001;39:277-303.
38. Tehranzadeh J, Wong E, Wang F, Sadighpour M. Imaging of osteomyelitis in the mature skeleton. *Radiologic clinics of North America* 2001;39:223-50.
39. Vukanovic S, Hauser H, Wettstein P. CT localization of myonecrosis for surgical decompression. *AJR Am J*

- Roentgenol 1980;135:1298.
40. Wysoki MG, Santora TA, Shah RM, Friedman AC. Necrotizing fasciitis: CT characteristics. *Radiology* 1997;203:859-63.
  41. Jacobson AF, Harley JD, Lipsky BA, Pecoraro RE. Diagnosis of osteomyelitis in the presence of soft-tissue infection and radiologic evidence of osseous abnormalities: value of leukocyte scintigraphy. *AJR Am J Roentgenol* 1991;157:807-12.
  42. Johnson JE, Kennedy EJ, Shereff MJ, Patel NC, Collier BD. Prospective study of bone, indium-111-labeled white blood cell, and gallium-67 scanning for the evaluation of osteomyelitis in the diabetic foot. *Foot & ankle international* 1996;17:10-6.
  43. Kolindou A, Liu Y, Ozker K, et al. In-111 WBC imaging of osteomyelitis in patients with underlying bone scan abnormalities. *Clin Nucl Med* 1996;21:183-91.
  44. Palestro CJ, Love C, Bhargava KK. Labeled leukocyte imaging: current status and future directions. *Q J Nucl Med Mol Imaging* 2009;53:105-23.
  45. Palestro CJ, Roumanas P, Swyer AJ, Kim CK, Goldsmith SJ. Diagnosis of musculoskeletal infection using combined In-111 labeled leukocyte and Tc-99m SC marrow imaging. *Clin Nucl Med* 1992;17:269-73.
  46. Wolf G, Aigner RM, Schwarz T. Diagnosis of bone infection using 99m Tc-HMPAO labelled leukocytes. *Nucl Med Commun* 2001;22:1201-6.
  47. Joseph TN, Mujtaba M, Chen AL, et al. Efficacy of combined technetium-99m sulfur colloid/indium-111 leukocyte scans to detect infected total hip and knee arthroplasties. *J Arthroplasty* 2001;16:753-8.
  48. Magnuson JE, Brown ML, Hauser MF, Berquist TH, Fitzgerald RH, Jr., Klee GG. In-111-labeled leukocyte scintigraphy in suspected orthopedic prosthesis infection: comparison with other imaging modalities. *Radiology* 1988;168:235-9.
  49. Mandell GA, Contreras SJ, Conard K, Harcke HT, Maas KW. Bone scintigraphy in the detection of chronic recurrent multifocal osteomyelitis. *J Nucl Med* 1998;39:1778-83.
  50. Palestro CJ. Radionuclide imaging of osteomyelitis. *Seminars in nuclear medicine* 2015;45:32-46.
  51. Thang SP, Tong AK, Lam WW, Ng DC. SPECT/CT in musculoskeletal infections. *Semin Musculoskelet Radiol* 2014;18:194-202.
  52. Buhne KH, Bohndorf K. Imaging of posttraumatic osteomyelitis. *Semin Musculoskelet Radiol* 2004;8:199-204.
  53. Crymes WB, Jr., Demos H, Gordon L. Detection of musculoskeletal infection with 18F-FDG PET: review of the current literature. *J Nucl Med Technol* 2004;32:12-5.
  54. De Winter F, Vogelaers D, Gemmel F, Dierckx RA. Promising role of 18-F-fluoro-D-deoxyglucose positron emission tomography in clinical infectious diseases. *Eur J Clin Microbiol Infect Dis* 2002;21:247-57.
  55. Guhlmann A, Brecht-Krauss D, Suger G, et al. Chronic osteomyelitis: detection with FDG PET and correlation with histopathologic findings. *Radiology* 1998;206:749-54.
  56. Mettler FA, Guiberteau MJ. Inflammation and infection imaging. *Essentials of Nuclear Medicine Imaging*. 5th ed. Philadelphia, Pa: Saunders Elsevier; 2006:341-58.
  57. Sugawara Y, Braun DK, Kison PV, Russo JE, Zasadny KR, Wahl RL. Rapid detection of human infections with fluorine-18 fluorodeoxyglucose and positron emission tomography: preliminary results. *Eur J Nucl Med* 1998;25:1238-43.
  58. Schiesser M, Stumpe KD, Trentz O, Kossmann T, Von Schulthess GK. Detection of metallic implant-associated infections with FDG PET in patients with trauma: correlation with microbiologic results. *Radiology* 2003;226:391-8.
  59. Stumpe KD, Zanetti M, Weishaupt D, Hodler J, Boos N, Von Schulthess GK. FDG positron emission tomography for differentiation of degenerative and infectious endplate abnormalities in the lumbar spine detected on MR imaging. *AJR Am J Roentgenol* 2002;179:1151-7.
  60. Ledermann HP, Schweitzer ME, Morrison WB. Nonenhancing tissue on MR imaging of pedal infection: characterization of necrotic tissue and associated limitations for diagnosis of osteomyelitis and abscess. *AJR Am J Roentgenol* 2002;178:215-22.
  61. Zacharias N, Velmahos GC, Salama A, et al. Diagnosis of necrotizing soft tissue infections by computed tomography. *Arch Surg* 2010;145:452-5.
  62. Deininger-Czermak E, Heimer J, Tappero C, Thali MJ, Gascho D. The added value of postmortem magnetic resonance imaging in cases of hanging compared to postmortem computed tomography and autopsy. *Forensic Sci Med Pathol* 2020;16:234-42.

63. Hayeri MR, Ziai P, Shehata ML, Teytelboym OM, Huang BK. Soft-Tissue Infections and Their Imaging Mimics: From Cellulitis to Necrotizing Fasciitis. *Radiographics* 2016;36:1888-910.
64. Yoon MA, Chung HW, Yeo Y, et al. Distinguishing necrotizing from non-necrotizing fasciitis: a new predictive scoring integrating MRI in the LRINEC score. *Eur Radiol* 2019;29:3414-23.
65. Fernando SM, Tran A, Cheng W, et al. Necrotizing Soft Tissue Infection: Diagnostic Accuracy of Physical Examination, Imaging, and LRINEC Score: A Systematic Review and Meta-Analysis. *Ann Surg* 2019;269:58-65.
66. Talbot BS, Weinberg EP. MR Imaging with Metal-suppression Sequences for Evaluation of Total Joint Arthroplasty. *Radiographics* 2016;36:209-25.
67. Khodarahmi I, Isaac A, Fishman EK, Dalili D, Fritz J. Metal About the Hip and Artifact Reduction Techniques: From Basic Concepts to Advanced Imaging. *Semin Musculoskelet Radiol* 2019;23:e68-e81.
68. Buckwalter KA. Optimizing imaging techniques in the postoperative patient. *Semin Musculoskelet Radiol* 2007;11:261-72.
69. Love C, Marwin SE, Palestro CJ. Nuclear medicine and the infected joint replacement. *Seminars in nuclear medicine* 2009;39:66-78.
70. Sofka CM, Potter HG, Adler RS, Pavlov H. Musculoskeletal imaging update: current applications of advanced imaging techniques to evaluate the early and long-term complications of patients with orthopedic implants. *HSS J* 2006;2:73-7.
71. White LM, Buckwalter KA. Technical considerations: CT and MR imaging in the postoperative orthopedic patient. *Semin Musculoskelet Radiol* 2002;6:5-17.
72. Chihara S, Segreti J. Osteomyelitis. *Dis Mon* 2010;56:5-31.
73. Wyers MR, Samet JD, Mithal LB. Physeal separation in pediatric osteomyelitis. *Pediatr Radiol* 2019;49:1229-33.
74. Towers JD. The use of intravenous contrast in MRI of extremity infection. *Semin Ultrasound CT MR* 1997;18:269-75.
75. Yu JS, Habib P. MR imaging of urgent inflammatory and infectious conditions affecting the soft tissues of the musculoskeletal system. *Emerg Radiol* 2009;16:267-76.
76. Edlich RF, Cross CL, Dahlstrom JJ, Long WB, 3rd. Modern concepts of the diagnosis and treatment of necrotizing fasciitis. *J Emerg Med* 2010;39:261-5.
77. Chun CW, Jung JY, Baik JS, Jee WH, Kim SK, Shin SH. Detection of soft-tissue abscess: Comparison of diffusion-weighted imaging to contrast-enhanced MRI. *J Magn Reson Imaging* 2018;47:60-68.
78. Reinus WR, De Cotiis D, Schaffer A. Changing patterns of septic tenosynovitis of the distal extremities. *Emerg Radiol* 2015;22:133-9.
79. Theodorou SJ, Theodorou DJ, Resnick D. Imaging findings of complications affecting the upper extremity in intravenous drug users: featured cases. *Emerg Radiol* 2008;15:227-39.
80. American College of Radiology. ACR practice parameter for performing and interpreting magnetic resonance imaging (MRI). Available at: <https://www.acr.org/-/media/ACR/Files/Practice-Parameters/MR-Perf-Interpret.pdf>. Accessed January 14, 2020.
81. American College of Radiology. ACR guidance document on MR safe practices: 2020. Available at: <https://www.acr.org/-/media/ACR/Files/Radiology-Safety/MR-Safety/Manual-on-MR-Safety.pdf>. Accessed July 6, 2020.
82. Shellock FG. *Reference Manual for Magnetic Resonance Safety, Implants, and Devices*. 2009 ed. Los Angeles, Calif: Biomedical Research Publishing Group; 2009.
83. Shellock FG, Crues JV. MR procedures: biologic effects, safety, and patient care. *Radiology* 2004;232:635-52.
84. American College of Radiology. ACR–SPR practice parameter for the use of intravascular contrast media. Available at: <https://www.acr.org/-/media/ACR/Files/Practice-Parameters/IVCM.pdf>. Accessed January 14, 2020.
85. American College of Radiology. ACR–SIR practice parameter for sedation/analgesia. Available at: <https://www.acr.org/-/media/ACR/Files/Practice-Parameters/Sed-Analgesia.pdf>. Accessed January 14, 2020.
86. Hovi I, Valtonen M, Korhola O, Hekali P. Low-field MR imaging for the assessment of therapy response in musculoskeletal infections. *Acta Radiol* 1995;36:220-7.
87. Miller TT, Randolph DA, Jr., Staron RB, Feldman F, Cushin S. Fat-suppressed MRI of musculoskeletal infection: fast T2-weighted techniques versus gadolinium-enhanced T1-weighted images. *Skeletal Radiol*

- 1997;26:654-8.
88. Potter HG, Nestor BJ, Sofka CM, Ho ST, Peters LE, Salvati EA. Magnetic resonance imaging after total hip arthroplasty: evaluation of periprosthetic soft tissue. *J Bone Joint Surg Am* 2004;86-A:1947-54.
  89. Ramnath RR. 3T MR imaging of the musculoskeletal system (Part I): considerations, coils, and challenges. *Magnetic resonance imaging clinics of North America* 2006;14:27-40.
  90. Ramnath RR. 3T MR imaging of the musculoskeletal system (Part II): clinical applications. *Magnetic resonance imaging clinics of North America* 2006;14:41-62.
  91. Tang JS, Gold RH, Bassett LW, Seeger LL. Musculoskeletal infection of the extremities: evaluation with MR imaging. *Radiology* 1988;166:205-9.
  92. White LM, Kim JK, Mehta M, et al. Complications of total hip arthroplasty: MR imaging-initial experience. *Radiology* 2000;215:254-62.
  93. Firbank MJ, Harrison RM, Williams ED, Coulthard A. Quality assurance for MRI: practical experience. *The British journal of radiology* 2000;73:376-83.
  94. Kneeland JB, Shimakawa A, Wehrli FW. Effect of intersection spacing on MR image contrast and study time. *Radiology* 1986;158:819-22.
  95. Rubin DA, Kneeland JB. MR imaging of the musculoskeletal system: technical considerations for enhancing image quality and diagnostic yield. *AJR Am J Roentgenol* 1994;163:1155-63.
  96. Guillerman RP. Osteomyelitis and beyond. *Pediatr Radiol* 2013;43 Suppl 1:S193-203.
  97. Metwalli ZA, Kan JH, Munjal KA, Orth RC, Zhang W, Guillerman RP. MRI of suspected lower extremity musculoskeletal infection in the pediatric patient: how useful is bilateral imaging? *AJR Am J Roentgenol* 2013;201:427-32.
  98. Erickson SJ. High-resolution imaging of the musculoskeletal system. *Radiology* 1997;205:593-618.
  99. Delfaut EM, Beltran J, Johnson G, Rousseau J, Marchandise X, Cotten A. Fat suppression in MR imaging: techniques and pitfalls. *Radiographics* 1999;19:373-82.
  100. Mirowitz SA. Fast scanning and fat-suppression MR imaging of musculoskeletal disorders. *AJR Am J Roentgenol* 1993;161:1147-57.
  101. Rybicki FJ, Chung T, Reid J, Jaramillo D, Mulkern RV, Ma J. Fast three-point dixon MR imaging using low-resolution images for phase correction: a comparison with chemical shift selective fat suppression for pediatric musculoskeletal imaging. *AJR Am J Roentgenol* 2001;177:1019-23.
  102. Shuman WP, Baron RL, Peters MJ, Tazioli PK. Comparison of STIR and spin-echo MR imaging at 1.5 T in 90 lesions of the chest, liver, and pelvis. *AJR Am J Roentgenol* 1989;152:853-9.
  103. Ziedses des Plantes BG, Koster K. Comparison of low-field versus high-field MR imaging. *Eur J Radiol* 1995;20:156-8.
  104. Jaramillo D, Laor T. Pediatric musculoskeletal MRI: basic principles to optimize success. *Pediatr Radiol* 2008;38:379-91.
  105. Yao L, Pitts JT, Thomasson D. Isotropic 3D fast spin-echo with proton-density-like contrast: a comprehensive approach to musculoskeletal MRI. *AJR Am J Roentgenol* 2007;188:W199-201.
  106. Collins MS, Schaar MM, Wenger DE, Mandrekar JN. T1-weighted MRI characteristics of pedal osteomyelitis. *AJR Am J Roentgenol* 2005;185:386-93.
  107. Johnson PW, Collins MS, Wenger DE. Diagnostic utility of T1-weighted MRI characteristics in evaluation of osteomyelitis of the foot. *AJR Am J Roentgenol* 2009;192:96-100.
  108. McGuinness B, Wilson N, Doyle AJ. The "penumbra sign" on T1-weighted MRI for differentiating musculoskeletal infection from tumour. *Skeletal Radiol* 2007;36:417-21.
  109. Hanna SL, Langston JW, Gronemeyer SA, Fletcher BD. Subtraction technique for contrast-enhanced MR images of musculoskeletal tumors. *Magnetic resonance imaging* 1990;8:213-5.
  110. Smith RC, Constable RT, Reinhold C, McCauley T, Lange RC, McCarthy S. Fast spin echo STIR imaging. *J Comput Assist Tomogr* 1994;18:209-13.
  111. American College of Radiology. ACR manual on contrast media. Available at: <https://www.acr.org/Clinical-Resources/Contrast-Manual>. Accessed January 14, 2020.
  112. Gholamrezanezhad A, Basques K, Batouli A, Matcuk G, Alavi A, Jadvar H. Clinical Nononcologic Applications of PET/CT and PET/MRI in Musculoskeletal, Orthopedic, and Rheumatologic Imaging. *AJR Am J Roentgenol* 2018;210:W245-W63.
  113. Kim EY, Kwack KS, Cho JH, Lee DH, Yoon SH. Usefulness of dynamic contrast-enhanced MRI in differentiating between septic arthritis and transient synovitis in the hip joint. *AJR Am J Roentgenol*

- 2012;198:428-33.
114. Morelli JN, Runge VM, Ai F, et al. An image-based approach to understanding the physics of MR artifacts. *Radiographics* 2011;31:849-66.
  115. Kirchgessner T, Perlepe V, Michoux N, Larbi A, Vande Berg B. Fat suppression at 2D MR imaging of the hands: Dixon method versus CHESS technique and STIR sequence. *Eur J Radiol* 2017;89:40-46.
  116. Haacke EM, Lenz GW. Improving MR image quality in the presence of motion by using rephasing gradients. *AJR Am J Roentgenol* 1987;148:1251-8.
  117. Liu Y, Ji JX. Minimal-SAR RF pulse optimization for parallel transmission in MRI. *Conf Proc IEEE Eng Med Biol Soc* 2008;2008:5774-7.
  118. Alaia EF, Benedick A, Obuchowski NA, et al. Comparison of a fast 5-min knee MRI protocol with a standard knee MRI protocol: a multi-institutional multi-reader study. *Skeletal Radiol* 2018;47:107-16.
  119. Garwood ER, Recht MP, White LM. Advanced Imaging Techniques in the Knee: Benefits and Limitations of New Rapid Acquisition Strategies for Routine Knee MRI. *AJR Am J Roentgenol* 2017;209:552-60.
  120. Johnson PM, Recht MP, Knoll F. Improving the Speed of MRI with Artificial Intelligence. *Semin Musculoskelet Radiol* 2020;24:12-20.
  121. Peh WC, Chan JH. Artifacts in musculoskeletal magnetic resonance imaging: identification and correction. *Skeletal Radiol* 2001;30:179-91.
  122. Vandevenne JE, Vanhoenacker FM, Parizel PM, Butts Pauly K, Lang RK. Reduction of metal artefacts in musculoskeletal MR imaging. *JBR-BTR* 2007;90:345-9.
  123. Jiang MH, He C, Feng JM, et al. Magnetic resonance imaging parameter optimizations for diagnosis of periprosthetic infection and tumor recurrence in artificial joint replacement patients. *Sci Rep* 2016;6:36995.
  124. Muller GM, Lundin B, von Schewelov T, Muller MF, Ekberg O, Mansson S. Evaluation of metal artifacts in clinical MR images of patients with total hip arthroplasty using different metal artifact-reducing sequences. *Skeletal Radiol* 2015;44:353-9.
  125. Hayter CL, Koff MF, Shah P, Koch KM, Miller TT, Potter HG. MRI after arthroplasty: comparison of MAVRIC and conventional fast spin-echo techniques. *AJR Am J Roentgenol* 2011;197:W405-11.
  126. Jawhar A, Reichert M, Kostrzewa M, et al. Usefulness of slice encoding for metal artifact correction (SEMAC) technique for reducing metal artifacts after total knee arthroplasty. *Eur J Orthop Surg Traumatol* 2019;29:659-66.
  127. Liebl H, Heilmeyer U, Lee S, et al. In vitro assessment of knee MRI in the presence of metal implants comparing MAVRIC-SL and conventional fast spin echo sequences at 1.5 and 3 T field strength. *J Magn Reson Imaging* 2015;41:1291-9.
  128. Marshman LA, Strong G, Trehwella M, Kasis A, Friesem T. Minimizing ferromagnetic artefact with metallic lumbar total disc arthroplasty devices at adjacent segments: technical note. *Spine (Phila Pa 1976)* 2010;35:252-6.
  129. Yanasak NE, Kelly MJ. MR imaging artifacts and parallel imaging techniques with calibration scanning: a new twist on old problems. *Radiographics* 2014;34:532-48.
  130. Craig JG, Amin MB, Wu K, et al. Osteomyelitis of the diabetic foot: MR imaging-pathologic correlation. *Radiology* 1997;203:849-55.
  131. American College of Radiology. ACR practice parameter for communication of diagnostic imaging findings. Available at: <https://www.acr.org/-/media/ACR/Files/Practice-Parameters/CommunicationDiag.pdf>. Accessed January 14, 2020.
  132. Shellock FG. *Guide to MR Procedures and Metallic Objects: Update 2001*. 7th ed. Philadelphia, Pa: Lippincott Williams & Wilkins; 2001.
  133. Shellock FG, Spinazzi A. MRI safety update 2008: part 2, screening patients for MRI. *AJR Am J Roentgenol* 2008;191:1140-9.
  134. American College of Radiology. ACR–AAPM technical standard for diagnostic medical physics performance monitoring of magnetic resonance imaging (MRI) equipment. Available at: <https://www.acr.org/-/media/ACR/Files/Practice-Parameters/MR-Equip.pdf>. Accessed January 14, 2020.

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Revised 2021 (Resolution 43) This Practice Parameter

2011 (Resolution 22)

Amended 2014 (Resolution 39)

Revised 2016 (Resolution 4)

Revised 2021 (Resolution 43)

Amended 2023 (Resolution 2c)